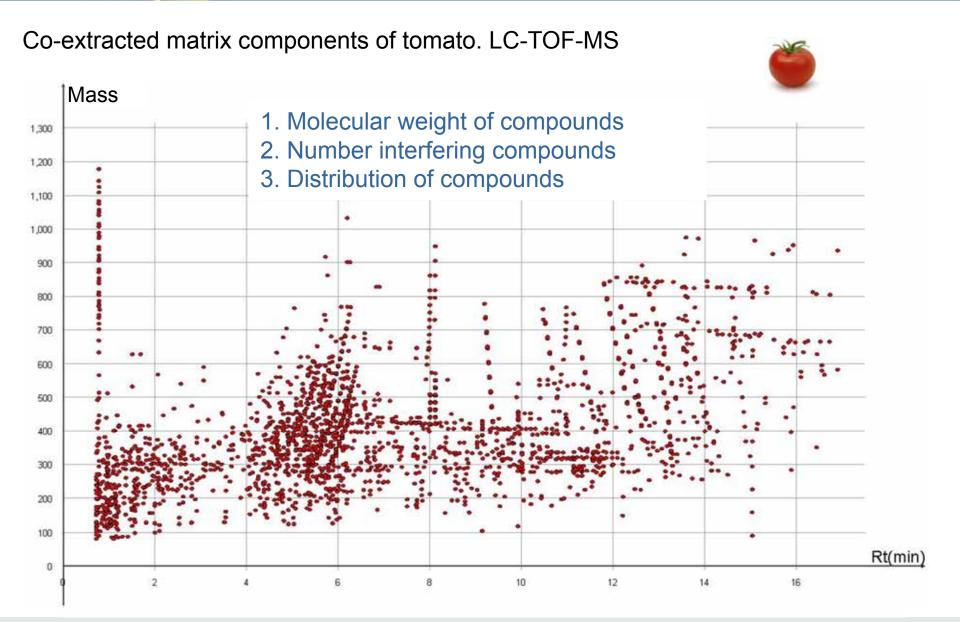
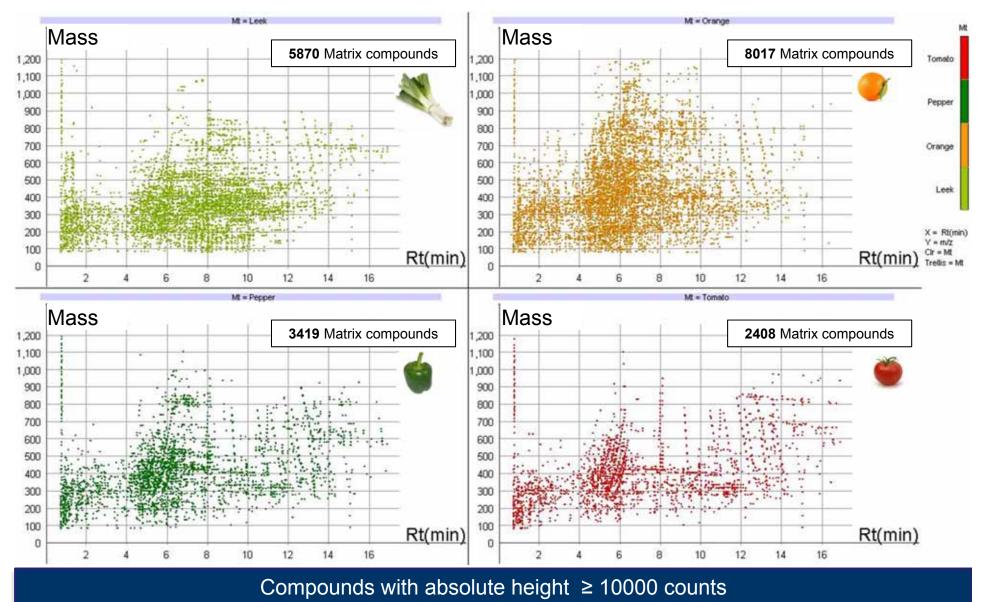


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Co-extracted matrix components LC-TOF-MS





LC-QqQ-MS/MS Skimer After 30 tea injections

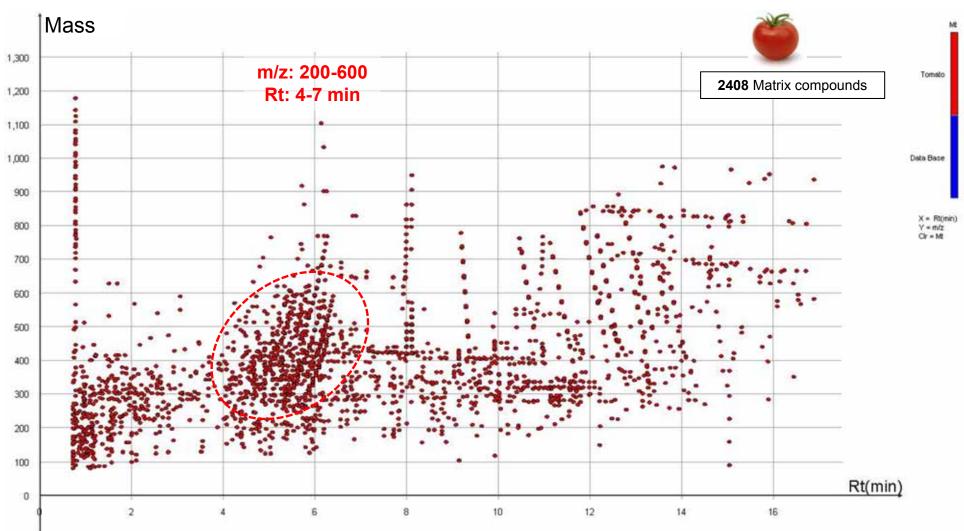


GC-Q-MS Liner after 40 tea injections



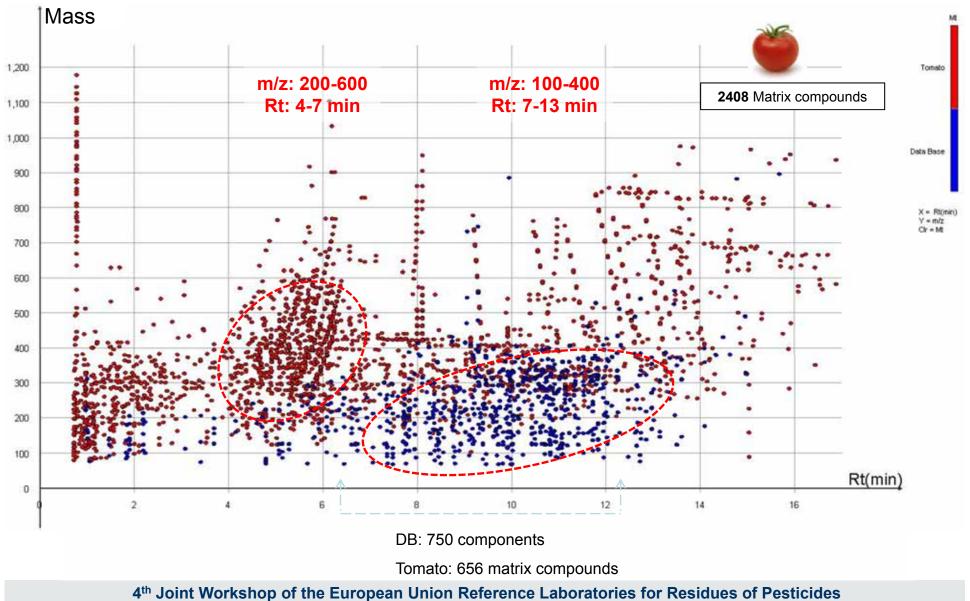


Co-extracted matrix components of tomato. LC-TOF-MS



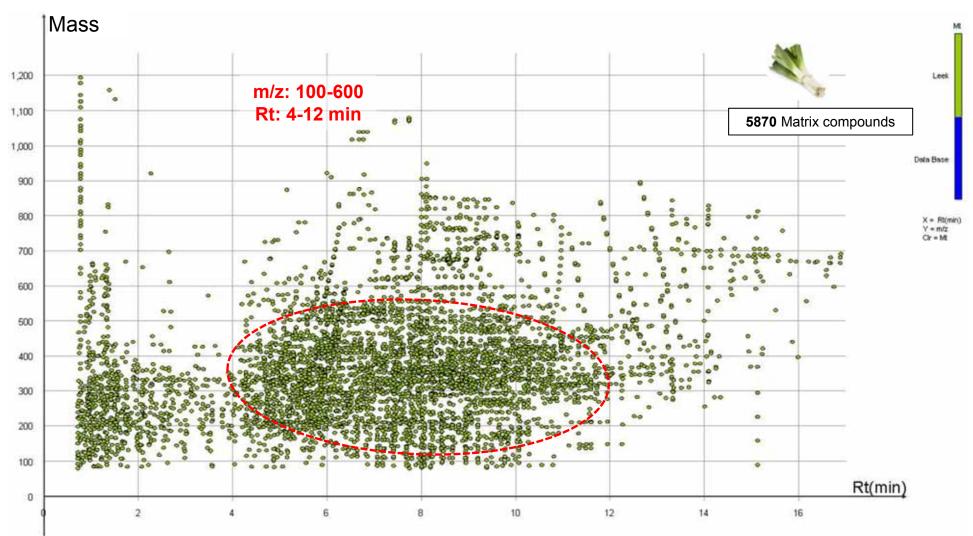


Data base components- tomato matrix compounds



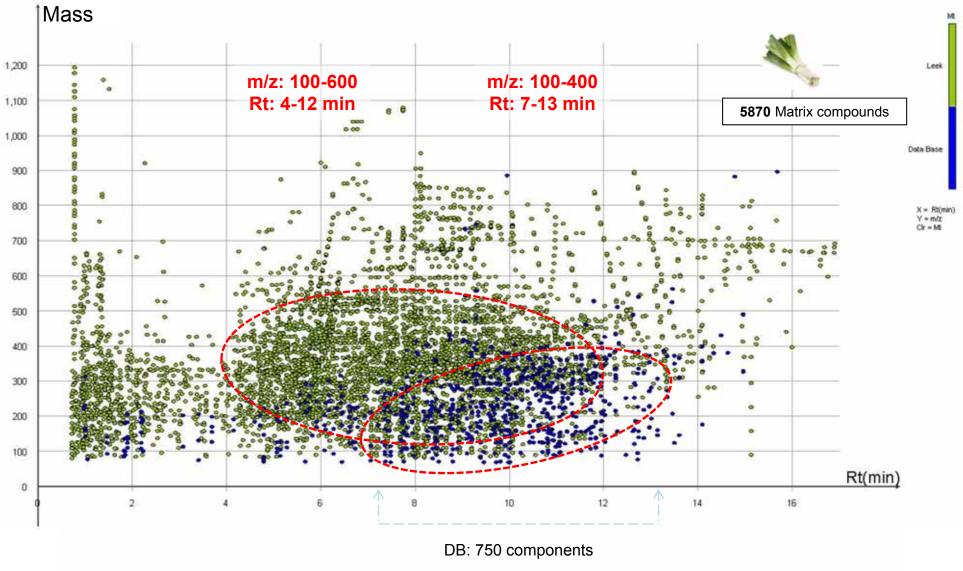


Co-extracted matrix components of leek. LC-TOF-MS





Data base components- leek matrix compounds



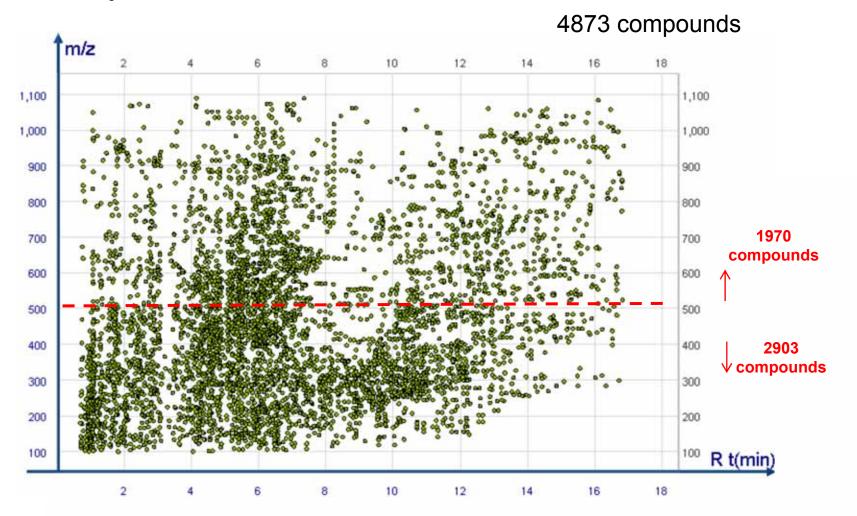
Leek: 3032 matrix compounds

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Matrix	N° of co-extracted compounds			
	Rt: 0-17 min	Rt: 7-13 min		
Green tea	9073	3535		
Orange	8017	2743		
Chamomille	6630	3490		
Leek	5870	3032		
Pepper	3419	919		
Tomato	2408	656		



Green tea matrix compounds Dil 1/5 . LC-TOF-MS Absolute height \geq 10000 counts

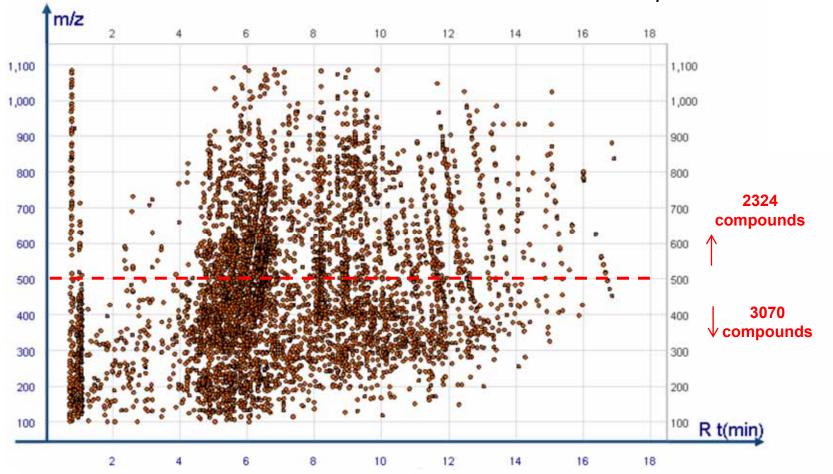




Orange matrix compounds Dil 1/5 . LC-TOF-MS

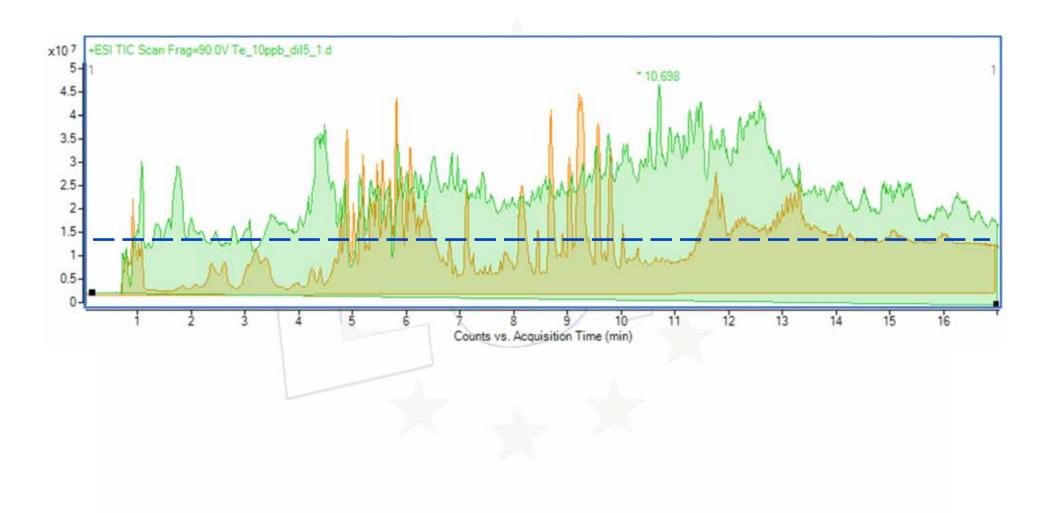
Absolute height \geq 10000 counts

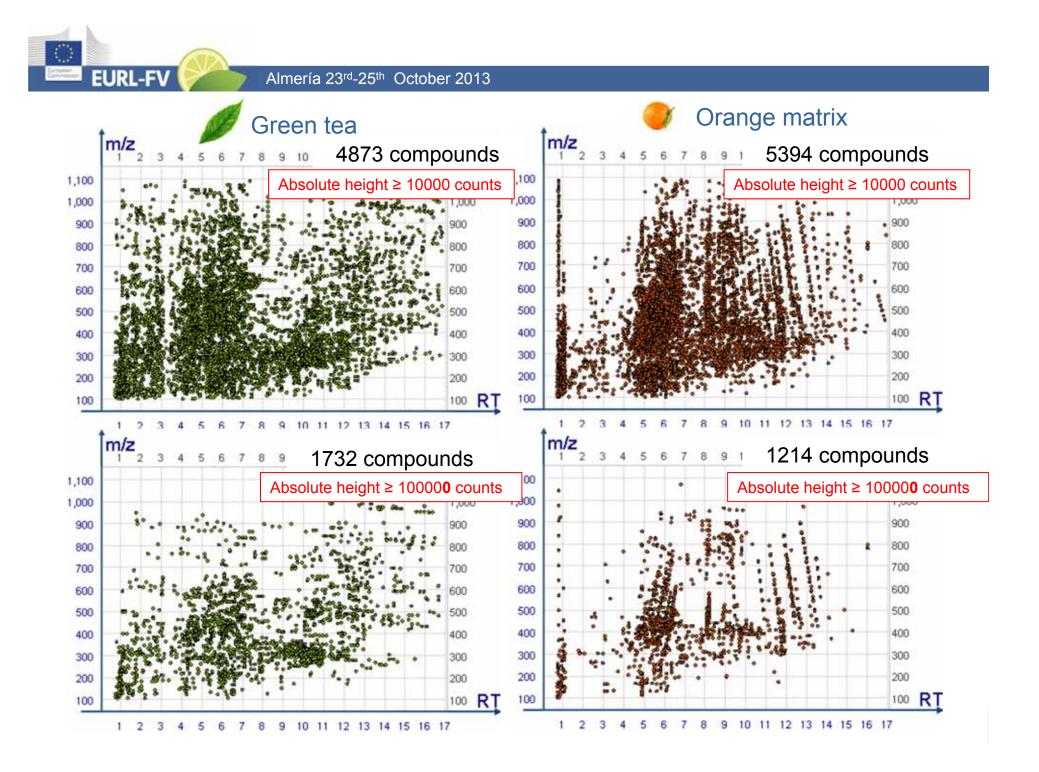
5394 compounds



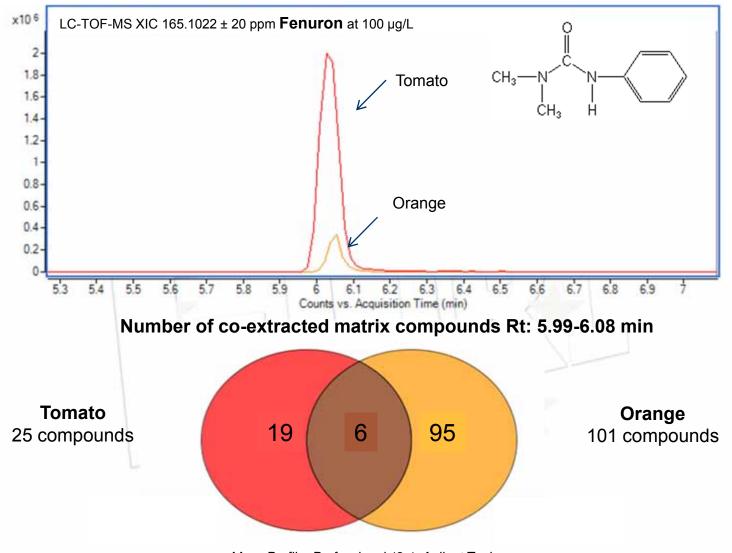


TIC LC-TOF-MS. Orange and Green tea. Dil 1/5

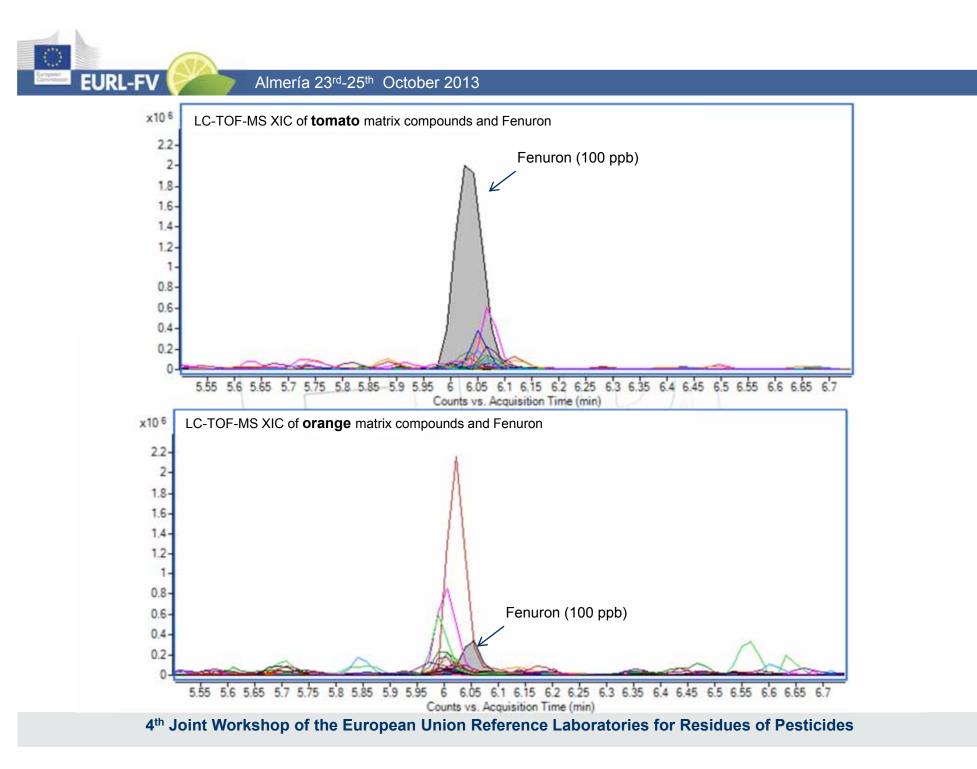




EURL-FV



Mass Profiler Professional 12.1. Agilent Tech.



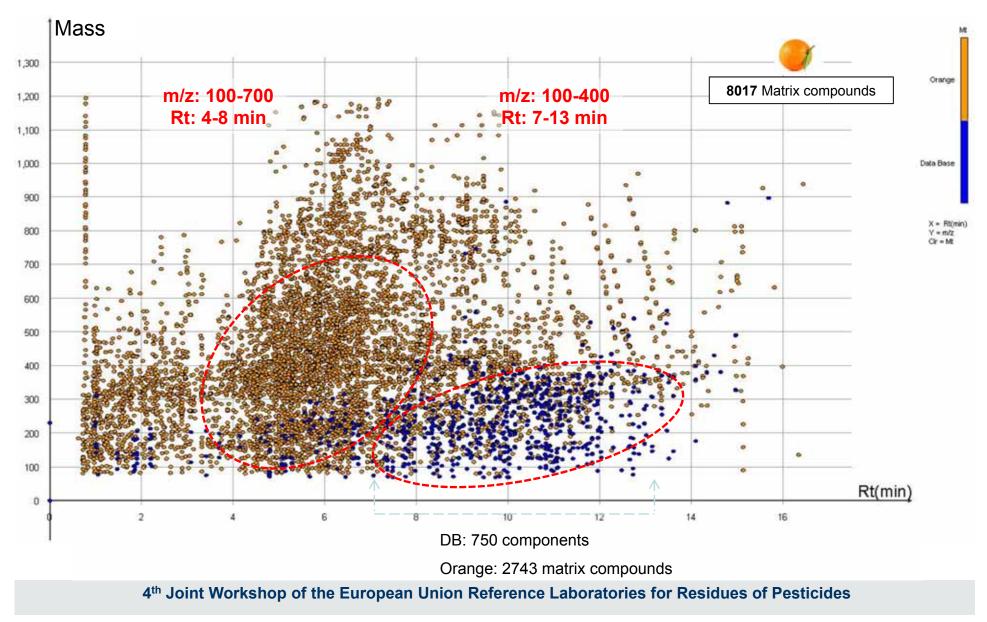
EURL-FV

Number of matrix coeluting compounds *vs* matrix suppression Orange matrix Dilution 1/30

Pesticide	Rt (min)	Suppression	N° of coleuting compounds (± 0.05 min)	Σ Compounds height (counts)
CINOSULFURON	8.09	-6	63	3.46E+06
FENPROPIDIN	8.16	-18	62	1.20E+07
PROPOXUR	8.23	0	45	4.12E+06
CARBOFURAN	8.32	0	34	3.20E+06
FORCHLORFENURON	8.54	-16	32	9.85E+05
FLUOMETURON	8.54	-11	33	1.05E+06
ISOPROTURON	8.72	-88	84	2.49E+07
METALAXYL	8.74	-74	84	2.49E+07
OFURACE	8.84	1	39	2.13E+06
HEPTENOPHOS	8.97	-8	65	6.86E+06
SPINOSYN A	9.02	-61	99	2.39E+07
METAZACHLOR	9.25	-83	93	3.06E+07
BUPIRIMATE	9.27	-83	125	4.00E+07
CYPRODINIL	9.35	-87	76	1.98E+07
TRIADIMENOL	9.42	-18	48	9.73E+06
FLAZASULFURON	9.42	-76	58	9.78E+06
PROMECARB	9.95	-15	42	2.68E+06
AZOXYSTROBIN	10.03	-4	29	5.33E+06

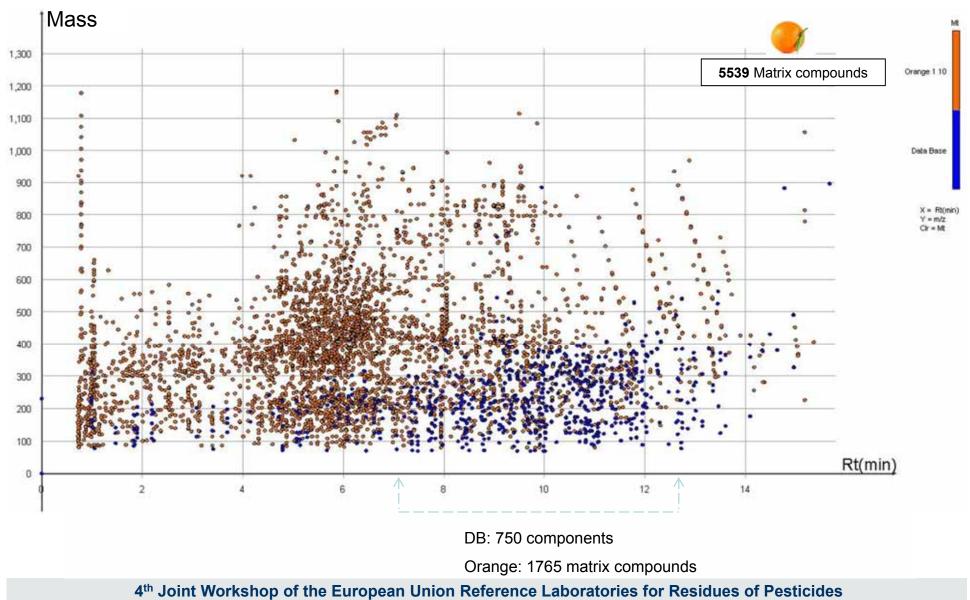


Data base components- orange matrix compounds





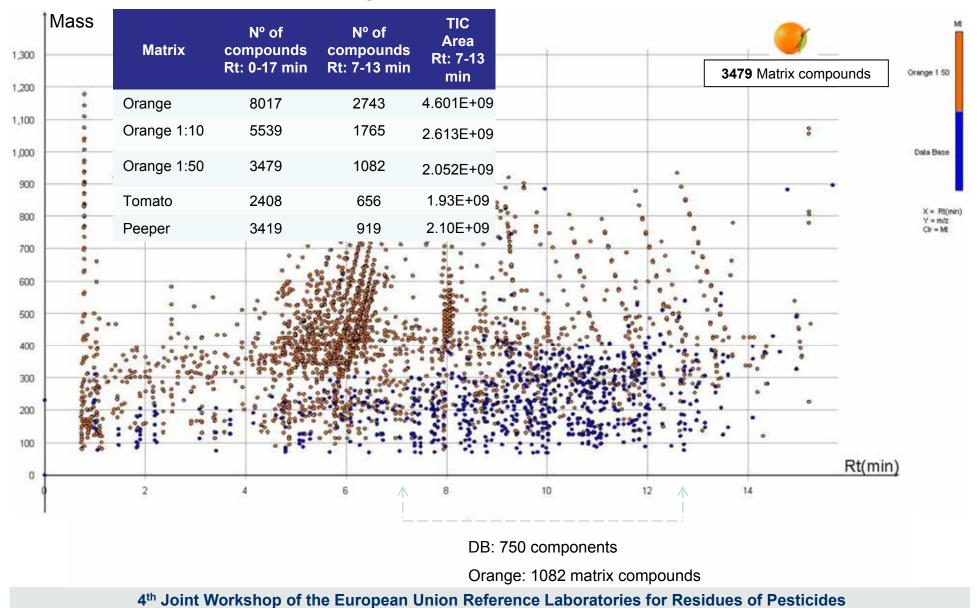
Data base components- orange matrix compounds. Dilution 1:10



EURL-FV

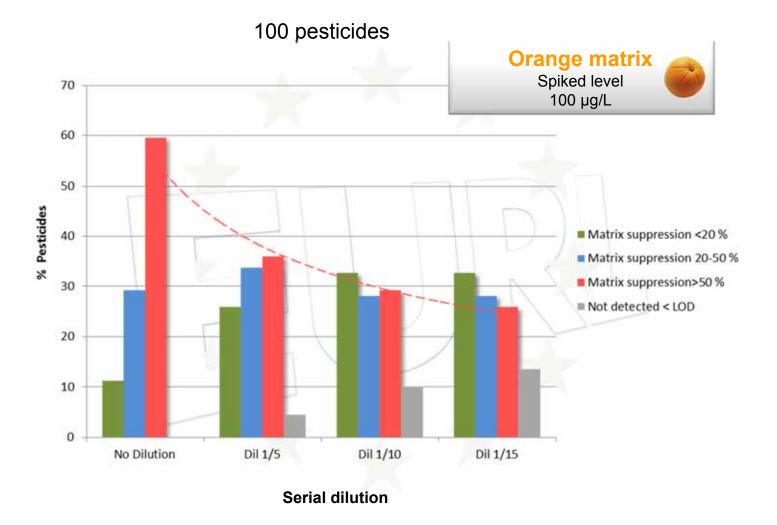
Almería 23rd-25th October 2013

Data base components- orange matrix compounds. Dilution 1:50





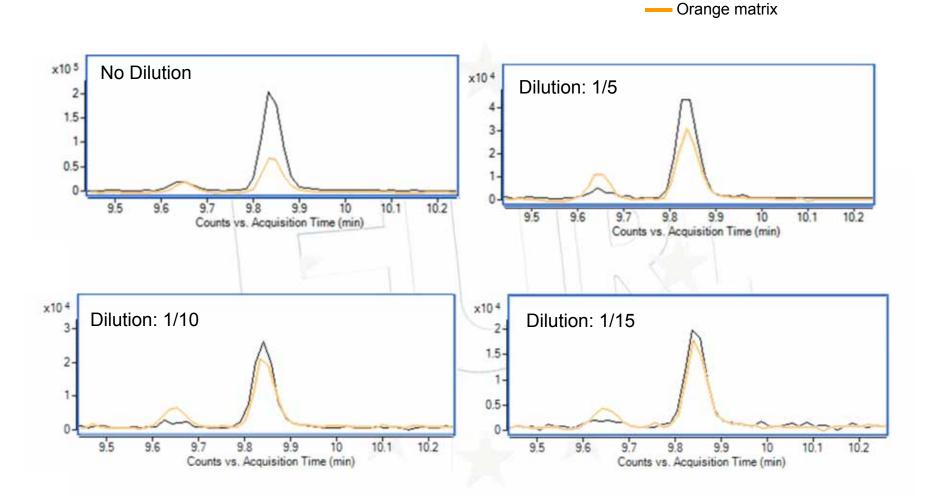
Dilution effect on matrix suppression LC-TOF-MS



Dilution effect on suppression decrease

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LC-TOF-MS XIC 268.1543 ± 20 ppm Diethofencarb at 100 µg/L

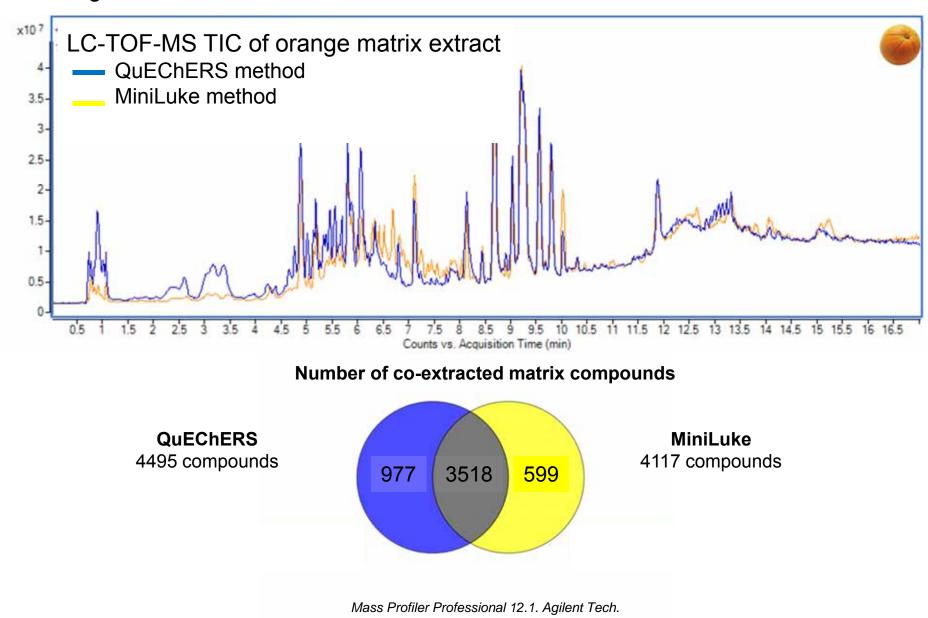


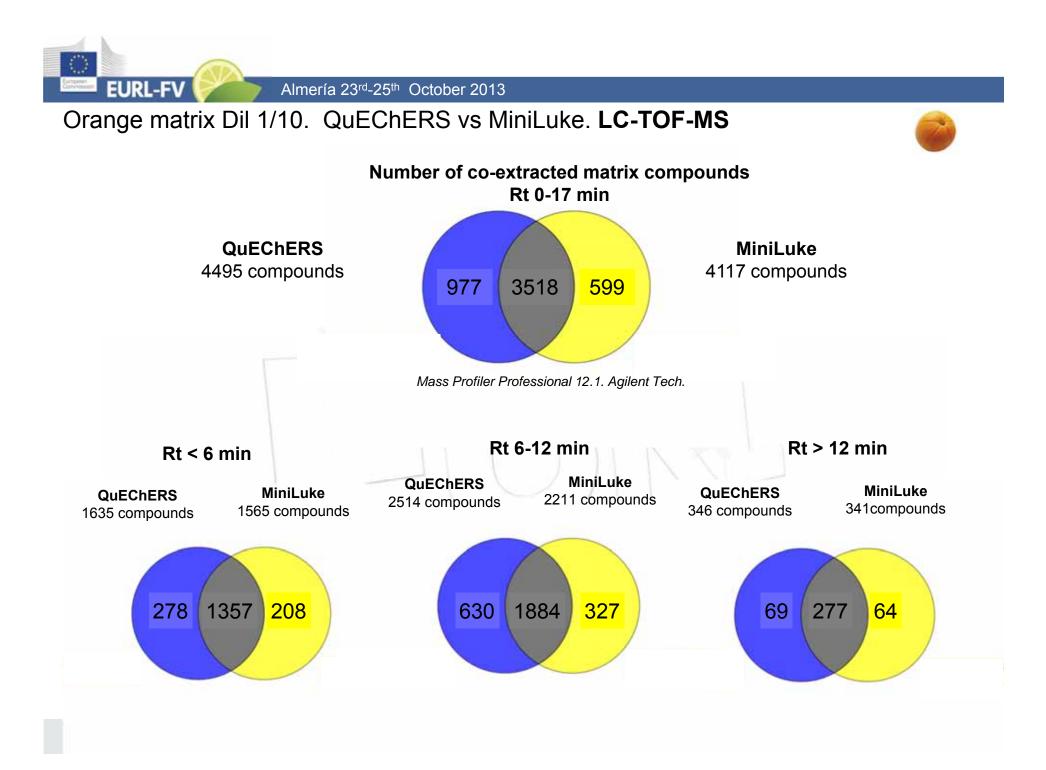
-Solvent

Orange matrix Dil 1/10. QuEChERS vs MiniLuke. LC-TOF-MS

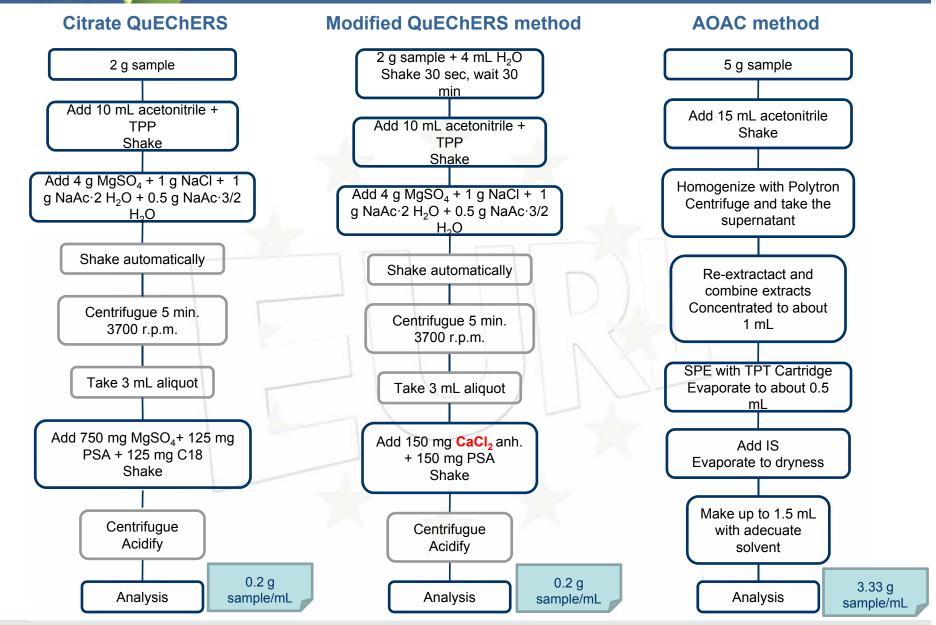
Almería 23rd-25th October 2013

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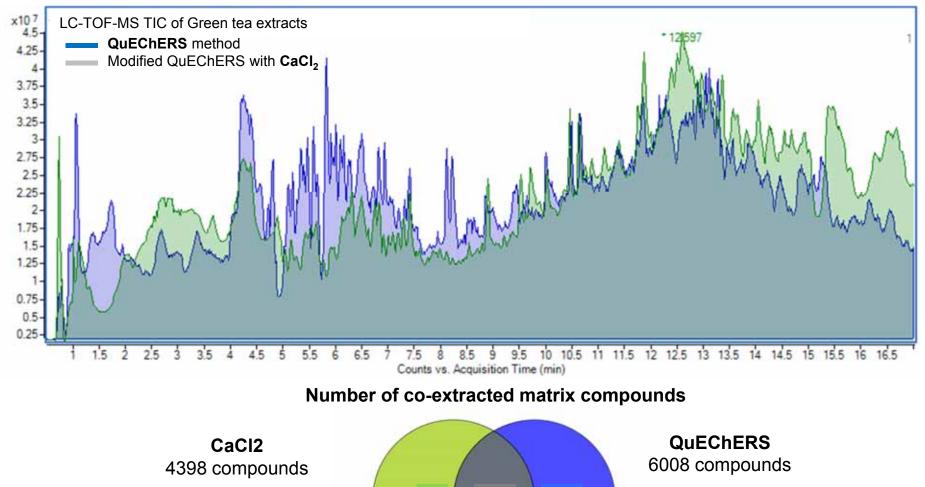


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Green tea extracts. CaCl2 vs QuEChERS method. LC-TOF-MS



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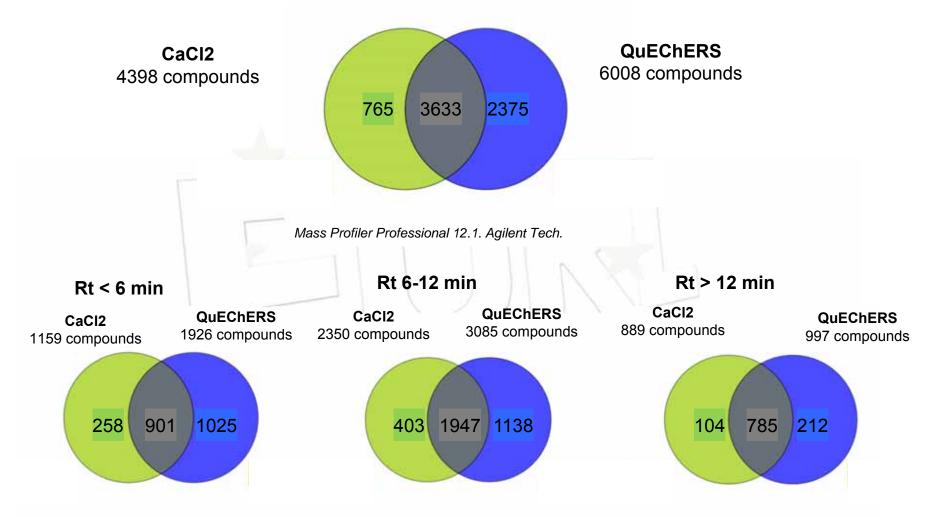
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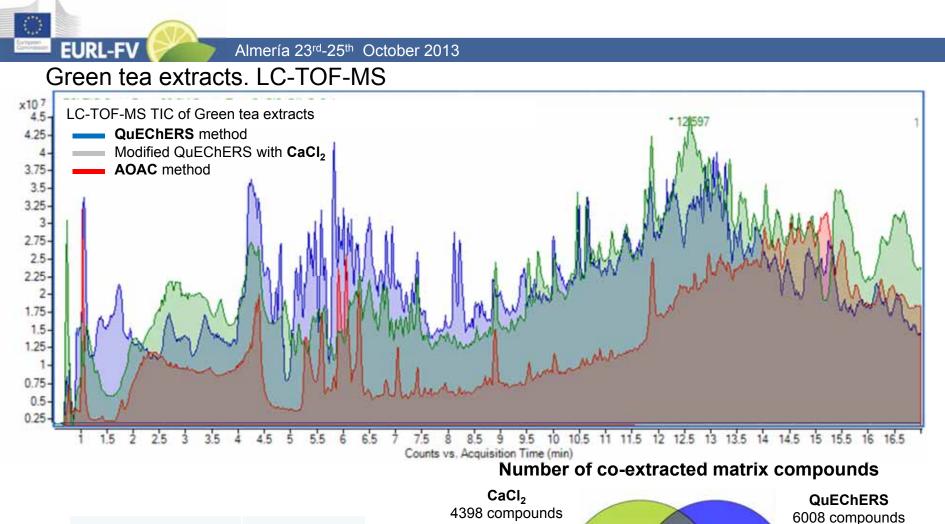
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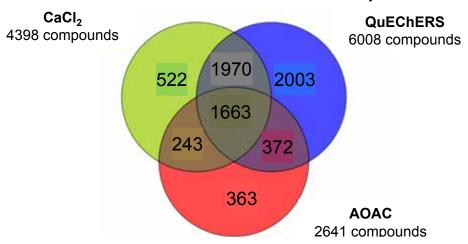
Green tea extracts. CaCl2 vs QuEChERS method. LC-TOF-MS

Number of co-extracted matrix compounds



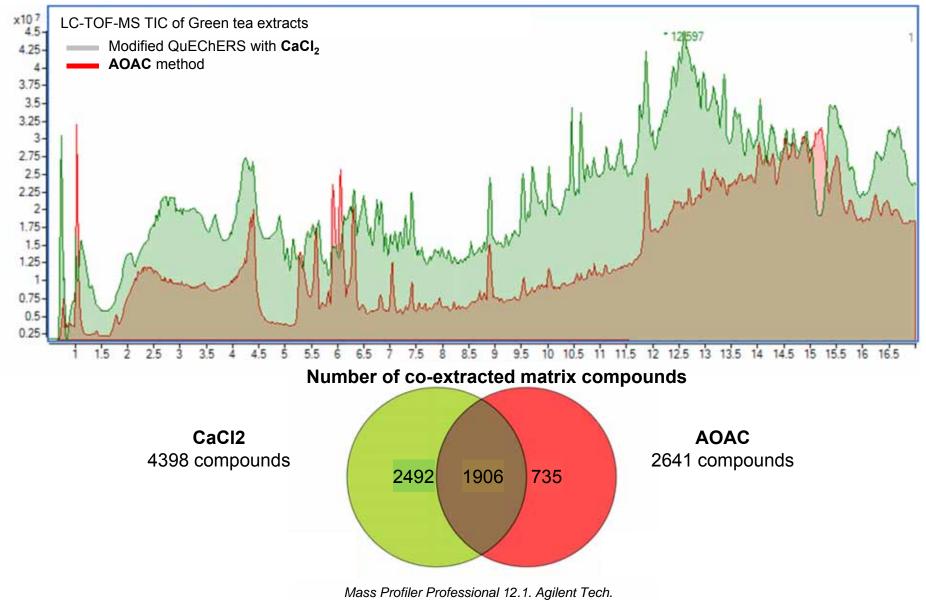


Methods	ΣArea
CaCl ₂	8.34E+09
AOAC	2.67E+09
QuEChERs	9.87E+09



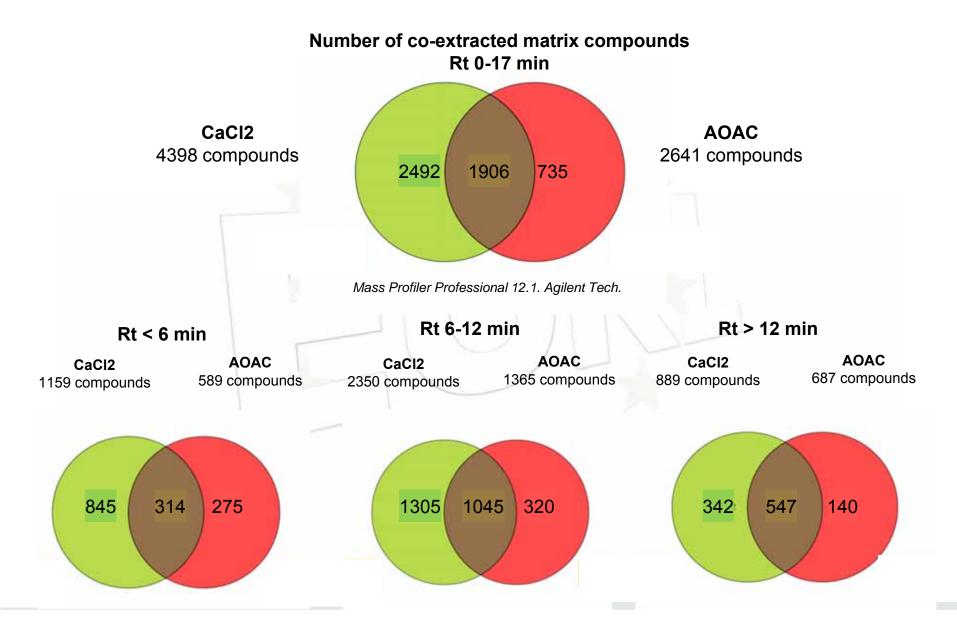
Green tea extracts. CaCl2 vs AOAC method. LC-TOF-MS

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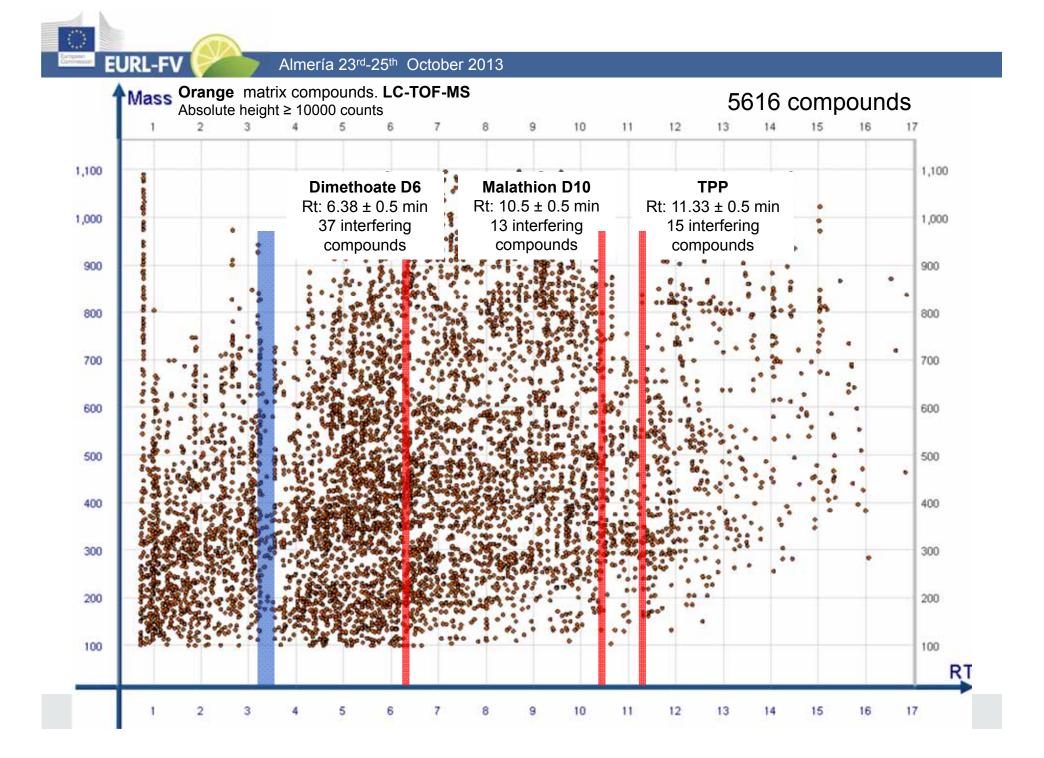


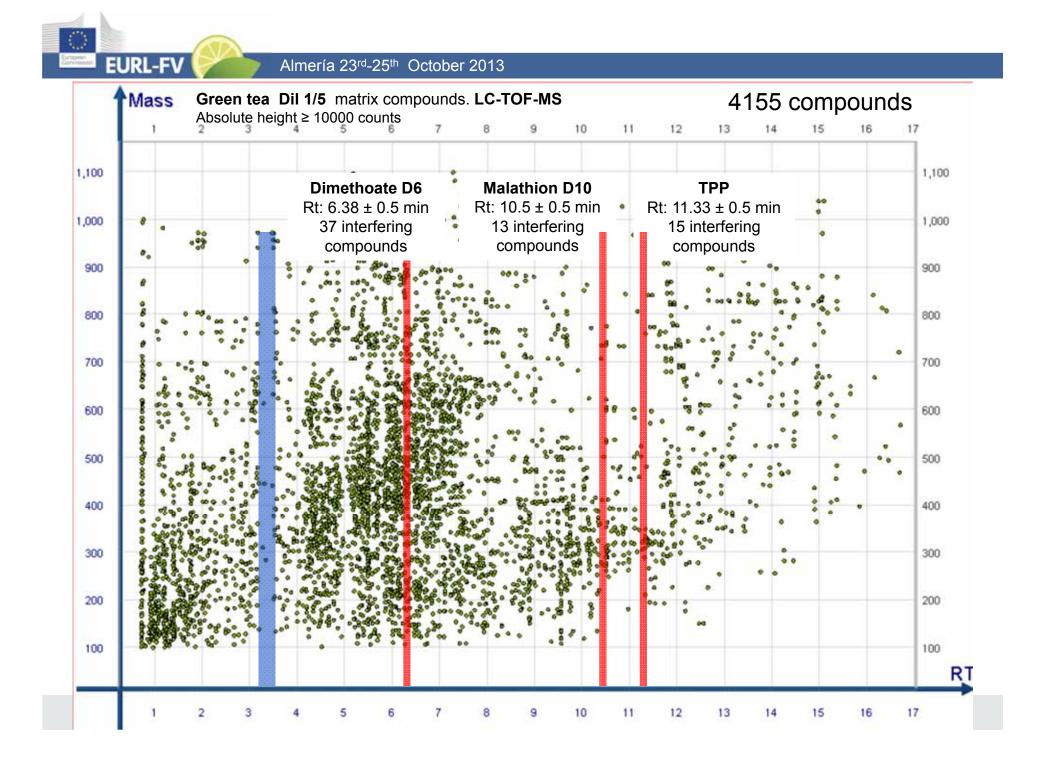
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Green tea extracts. CaCl2 vs AOAC method. LC-TOF-MS



EURL-FV Almería 23rd-25th October 2013 Tomato matrix compounds. LC-TOF-MS Mass 3304 compounds Absolute height \geq 10000 counts 1,100 1,100 TPP **Dimethoate D6** Malathion D10 Rt: 10.5 ± 0.5 min Rt: 11.33 ± 0.5 min Rt: 6.38 ± 0.5 min 1,000 1,000 37 interfering 13 interfering 15 interfering compounds compounds compounds . RT

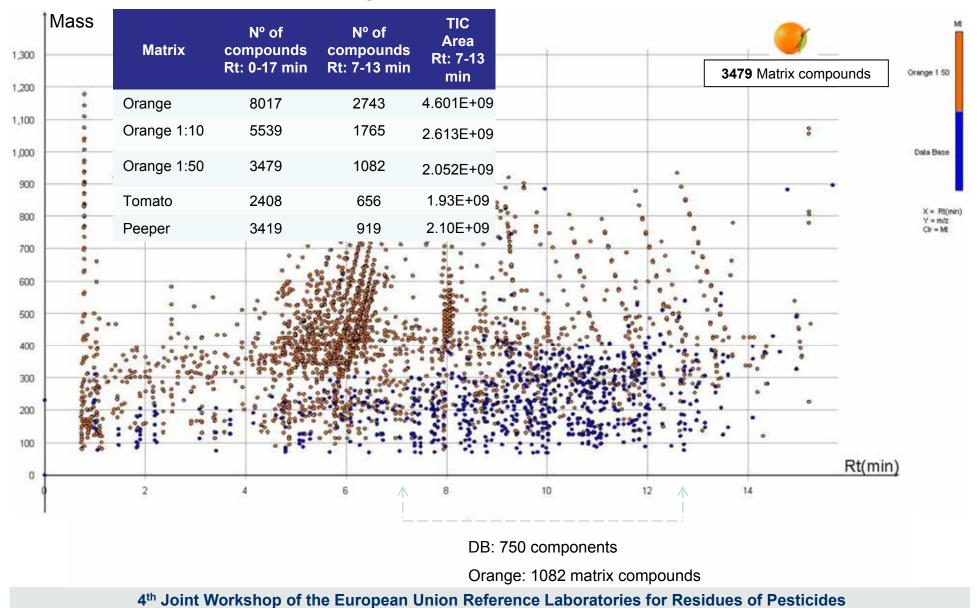


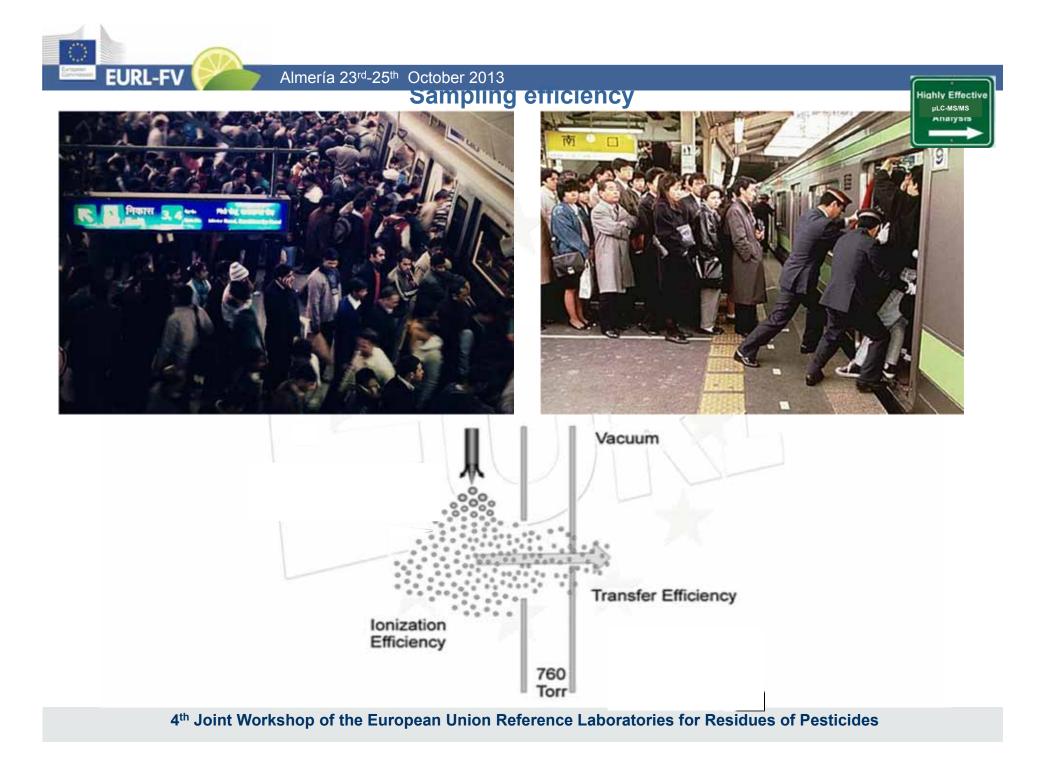


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Almería 23rd-25th October 2013

Data base components- orange matrix compounds. Dilution 1:50

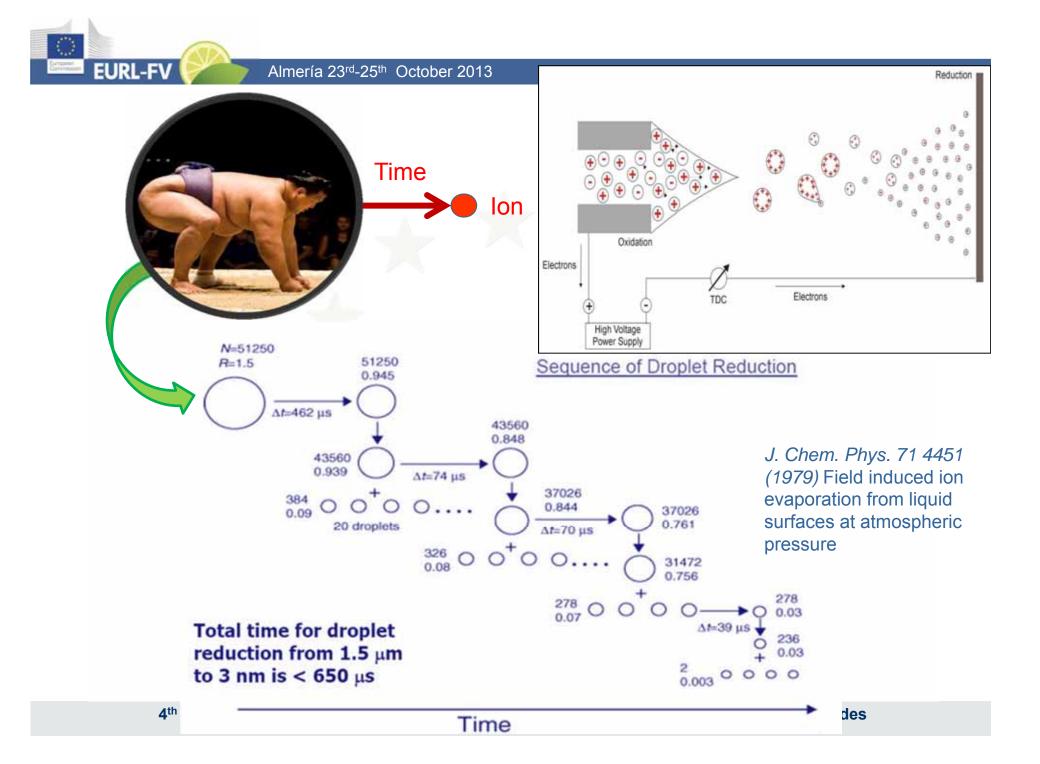








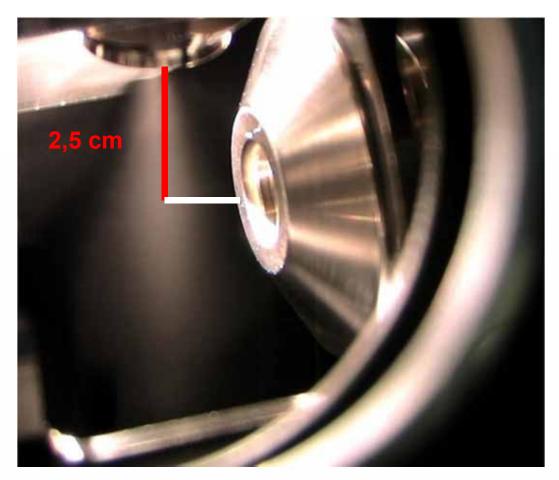
High temperature: 500°C Nitrogen flow: 40 L/min at 8 kg pressure Gas velocity: 100 – 300 m/sec (1000 km/h, almost Match1)







Observing Thermal Focusing



Temperature = 550 °C



Table 2 Gas flows generated with different GC and LC columns and solvent flows.

	Column	Carrier	Carrier flow-rate (ml min ⁻¹)	Gas flow atmosphere (ml min ⁻¹)	Gas flow (10 ⁻⁶ Torr) (×1000 l s ⁻¹)
	GC capillary	Helium	1	1	12.5
	GC packed/GC (CI)	Helium	20	20	250
L.	LC analytical	Hexane	1	184	2300
		MeOH	1	593	7400
		Water	1	1240	15500
	LC capillary	Hexane	0.010	1.8	23
		Water	0.010	12	155

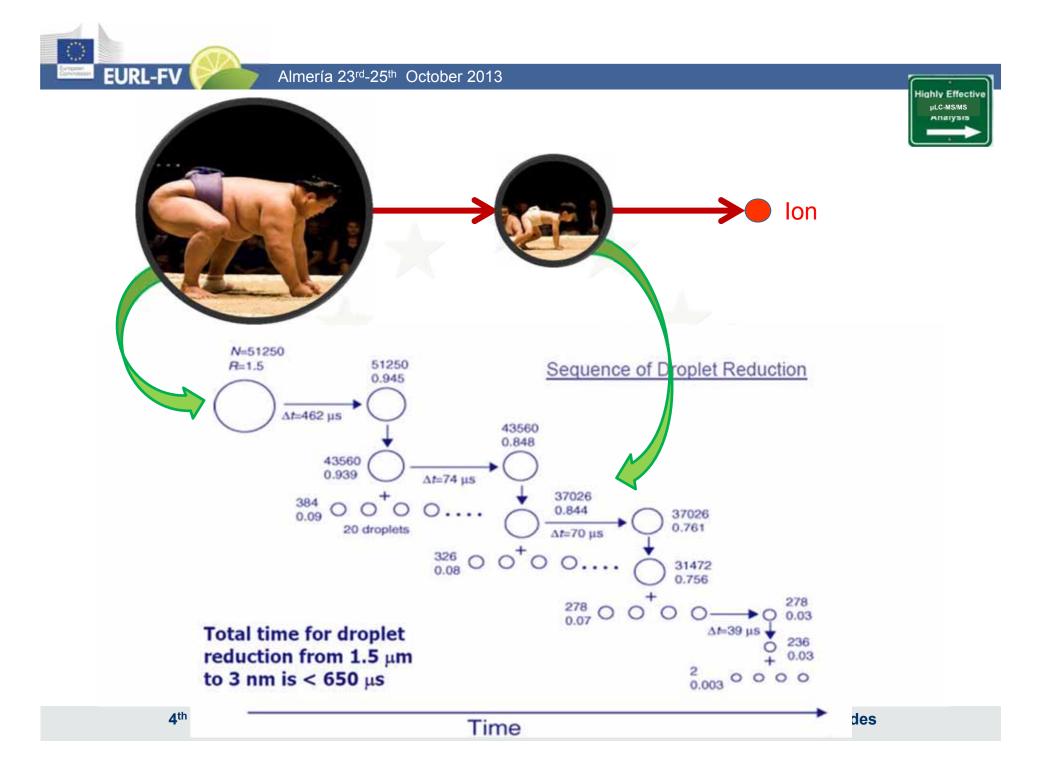
J. Abian et al. J. Mass Spectrom. 34, 157-168 (1999)

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Highly Effective µLC-MS/MS Analysis

Sampling efficiency

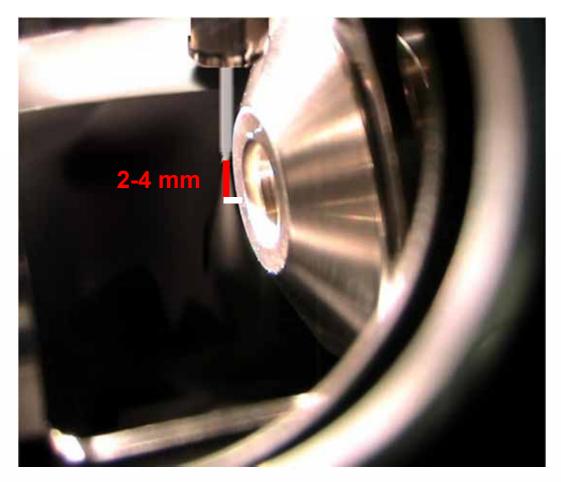








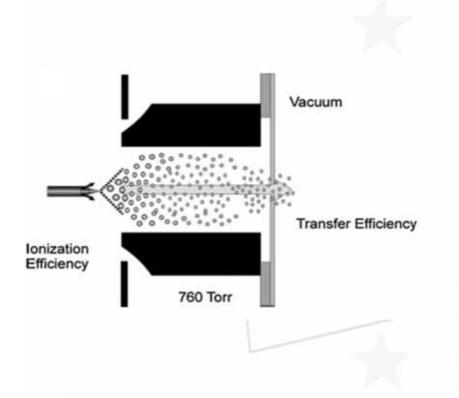
Observing Thermal Focusing



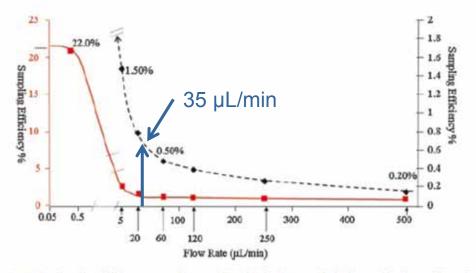
Temperature = 550 °C



Sampling efficiency



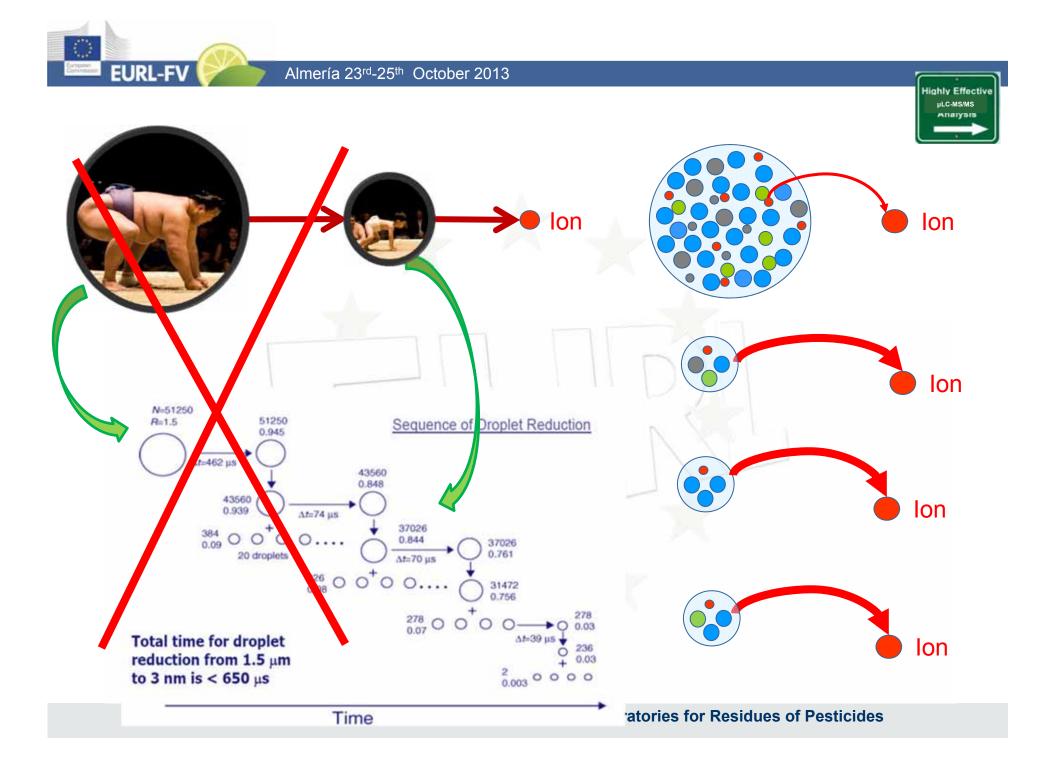
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Highly Effective µLC-MS/MS

FIGURE 9. Sampling efficiency versus flow rate. The data in the upper dashed trace (black), read from the right Yaxis, is an expansion of the data in the lower solid trace (red) read from the left Yaxis. For each flow the optimum source and interface configuration was used for those conditions. For high flow rates above 1 µL/min a heated TurboIonSpray[®] source with a conical pinhole aperture interface (standard interface) on an API 5000TM triple quadrupole instrument. For points below 1 µL/min a nanospray source with a 15 µm aperture fused silica capillary and an interface optimized for nanoflow introduction (PDI interface, See Ion Transport Section, Declustering, PDI interface) was used.

Atmospheric pressure ion sources. Thomas R. Covey *et al. Mass spectrometry Reviews*, **2009**, 28, 870-897



Standard-LC vs micro-LC

		Most used			l.	
	I.D. Column (mm)	Flow (µL/min)	Nebulizer	I.D. column (mm)	Flow (µL/min)	Sensitivity Theoretically Gain
Standard	9.0 1.6	200 -	Standard	4.6	400	1
LC	8.0 – 1.6	5000		2.1	200	5
	cro LC 0.3 – 1.0 5 - 200			1	40	20
micro LC		Micro	0.8	20	30	
				0.5	35	≈ 30



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Thanks for your Attention

